Species delimitation in endangered groundwater salamanders: Implications for aquifer management and biodiversity conservation

Thomas J. Devitt^{a,b,1,2}, April M. Wright^{a,b,3}, David C. Cannatella^{a,b}, and David M. Hillis^{a,b,1}

^aDepartment of Integrative Biology, The University of Texas at Austin, Austin, TX 78712; and ^bBiodiversity Center, The University of Texas at Austin, Austin, TX 78712

Contributed by David M. Hillis, November 20, 2018 (sent for review August 31, 2018; reviewed by Shawn R. Kuchta and C. Richard Tracy)

Groundwater-dependent species are among the least-known components of global biodiversity, as well as some of the most vulnerable because of rapid groundwater depletion at regional and global scales. The karstic Edwards-Trinity aquifer system of west-central Texas is one of the most species-rich groundwater systems in the world, represented by dozens of endemic groundwater-obligate species with narrow, naturally fragmented distributions. Here, we examine how geomorphological and hydrogeological processes have driven population divergence and speciation in a radiation of salamanders (Eurycea) endemic to the Edwards-Trinity system using phylogenetic and population genetic analysis of genome-wide DNA seguence data. Results revealed complex patterns of isolation and reconnection driven by surface and subsurface hydrology, resulting in both adaptive and nonadaptive population divergence and speciation. Our results uncover cryptic species diversity and refine the borders of several threatened and endangered species. The US Endangered Species Act has been used to bring state regulation to unrestricted groundwater withdrawals in the Edwards (Balcones Fault Zone) Aguifer, where listed species are found. However, the Trinity and Edwards-Trinity (Plateau) aquifers harbor additional species with similarly small ranges that currently receive no protection from regulatory programs designed to prevent groundwater depletion. Based on regional climate models that predict increased air temperature, together with hydrologic models that project decreased springflow, we conclude that Edwards-Trinity salamanders and other codistributed groundwater-dependent organisms are highly vulnerable to extinction within the next century.

endangered species | groundwater depletion | multispecies coalescent | phylogeography | species tree

roundwater is the largest source of liquid freshwater on Earth, providing drinking water for billions of people (1) and irrigation for most of the world's agriculture (2), while at the same time sustaining aquatic and terrestrial ecosystem function (3-5). Although groundwater was once a seemingly limitless resource that made possible the "green revolution" and expanded the global food supply (6-8), groundwater depletion has become an intensifying global problem (9-13). Aquifer overdraft threatens water and food security (14–16), as well as natural capital and economic welfare (17, 18). Groundwater-dependent ecosystems provide essential services (5), including water purification (19), biodegradation (20, 21), and nutrient cycling (22, 23). The groundwater-obligate organisms (stygobionts) underpinning these ecosystem services are among the least-known components of global biodiversity (24, 25), as well as some of the most vulnerable to extinction (see, e.g., refs. 26–28). They are particularly vulnerable because most have small distributions (25, 29) and are adapted to a narrow set of environmental conditions (3, 4, 30). The actual or functional extinction of narrowly endemic stygobionts may reverberate through ecological networks via complex interactions among species (see, e.g., refs. 31 and 32). In the worst case, extinction may result in the impaired functioning of groundwater-dependent aquatic and terrestrial ecosystems, resulting in the deterioration of water quality (see, e.g., refs. 33–35).

The karstic Edwards–Trinity aquifer system (36) of west-central Texas (Fig. 1) is one of the most species-rich groundwater environments in the world, represented by dozens of endemic species with small, naturally fragmented distributions (37–43). Groundwater is especially valuable for municipal, irrigation, and recreational use in this semiarid region, owing to low rainfall, frequent droughts, and little permanent surface water (44, 45). Groundwater development since the 1950s (46), combined with more recent, rapid residential development (47), has resulted in intensive pumping, decreased well yields, local water table declines, and diminished baseflow to springs and streams (48–51). Aguifer drawdown and the resulting reduction or cessation of springflow has resulted in habitat loss and fragmentation for groundwater species, compounded by reduced water quality from urban development (52–55). As a result, 13 groundwater-dependent species endemic to the Edwards Aquifer are listed as threatened or endangered under the US Endangered Species Act (ESA) (56-58),

Significance

Groundwater ecosystems deliver services that are vital to human well-being and environmental quality. Overexploitation of groundwater threatens biodiversity and ecosystem stability worldwide, underscoring the need to discover, describe, and sustain groundwater-dependent species before they are lost. Here we present a phylogenomic analysis of a salamander radiation endemic to the karstic Edwards-Trinity aquifer system of west-central Texas. This intensively used resource sustains narrow-range endemic groundwater species that are endangered with extinction due to aquifer overdraft. The federal Endangered Species Act has been used as a tool to bring regulation to groundwater withdrawal, but existing state law that treats groundwater as private property undermines conservation. Without ecologically relevant policies for sustainable use, unchecked groundwater depletion will likely result in species extinction and ecosystem degradation.

Author contributions: D.C.C. and D.M.H. designed research; T.J.D., A.M.W., and D.M.H. performed research; T.J.D., A.M.W., D.C.C., and D.M.H. analyzed data; and T.J.D. and D.M.H. wrote the paper.

Reviewers: S.R.K., Ohio University; and C.R.T., University of Nevada, Reno.

The authors declare no conflict of interest.

Published under the PNAS license.

Data deposition: Raw sequence reads have been deposited in the NCBI SRA database (accession no. PRJNA504898). Input files for all analyses have been deposited in the Dryad Digital Repository (doi: 10.5061/dryad.3k5c0sh).

See Commentary on page 2410.

¹To whom correspondence may be addressed. Email: tdevitt@utexas.edu or dhillis@austin.utexas.edu.

²Present address: Watershed Protection Department, City of Austin, Austin, TX 78704.

³Present address: Department of Biological Sciences, Southeastern Louisiana University, Hammond, LA 70402.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10. 1073/pnas.1815014116/-/DCSupplemental.

Published online January 14, 2019.



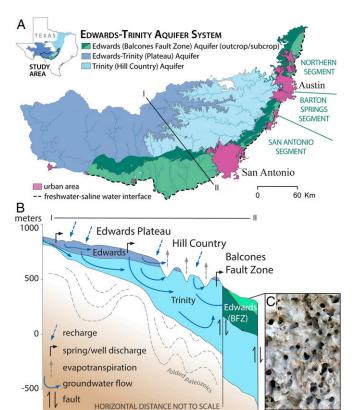


Fig. 1. (A) Map of the study area illustrating the extent of the Edwards-Trinity aguifer system in west-central Texas. (B) Conceptual hydrogeologic cross section of the Edwards-Trinity (Plateau), Trinity (Hill Country portion), and Edwards (BFZ) aquifers. Younger Edwards Group rocks overlie Trinity Group rocks except in the Hill Country where the Edwards formation has been mostly eroded. Cross section modified from ref. 86. (C) Photo illustrating highly porous Edwards Group limestone.

including a narrowly endemic mosquitofish (the San Marcos Gambusia, Gambusia georgei) that has not been seen since 1983 and is presumed extinct (59).

Among the federally listed aquatic species, over half are plethodontid salamanders of the genus Eurycea, members of a radiation of about 14 species (60-62) found in oligotrophic environments of the Edwards Aquifer and its catchment area in the Edwards-Trinity (Plateau) and Trinity (Hill Country) aguifers. These salamanders occur in surface springs and spring-fed streams, as well as below ground in water-filled caves, solutionally enlarged conduits, and interstices of the porous limestone rock matrix. Some species are only temporary inhabitants of the subsurface aquifer, whereas others are obligately subterranean; a few have both surface (epigean) and subsurface (hypogean) populations. Epigean species are generally morphologically similar overall (63– 65). Subterranean populations and species show phenotypic convergence (the independent evolution of similar traits) in their response to the challenges of life underground (62, 66–69). Previous genetic work using allozymes and mtDNA (60, 62) uncovered cryptic species (two or more morphologically similar species mistakenly classified as one; ref. 70) and revealed instances of shared ancestral polymorphism or introgressive hybridization that has made drawing species boundaries challenging. Species borders remain unclear for some taxa because many new populations have been discovered subsequent to the most recent taxonomic revision in 2000 (60). In some cases, newly discovered populations have been assigned to species only provisionally based on geographic proximity to known populations, without genetic evidence to support taxonomic designation (56). The accurate assignment of populations to species is essential in cases where the range size of a

taxon informs listing decisions about the vulnerability status of a species or the designation of critical habitat under the ESA (71, 72).

Because the species rank is the usual unit of currency for decision makers tasked with conservation prioritization, natural resource management, and environmental policy, accurately drawing species boundaries is critical for preventing the extinction of rare, cryptic species (see, e.g., ref. 73), spatial conservation prioritization (25), and ESA listing decisions (74). In this study, we use genome-wide DNA sequence data collected from over 300 individual Edwards-Trinity Eurycea salamanders to delimit evolutionary lineages at the species level and below. Our results uncover cryptic species and redraw the boundaries of some named species, including several that are listed under the ESA. These results have major implications for regional natural resource management and conservation because the ESA has been used as a tool to bring regulation to unrestricted groundwater withdrawal in the eastern half of the Edwards Aquifer where listed species are found (75–77). We also show that the Trinity and Edwards-Trinity (Plateau) aquifers harbor additional unnamed species that receive no protection, highlighting watersheds that deserve priority for biodiversity conservation.

The Edwards-Trinity Aquifer System

Three major laterally adjacent and stratigraphically layered aquifers make up the Edwards-Trinity system, coincident with geographic subregions of the Edwards Plateau defined by distinct physiography, hydrology, and geology (Fig. 1A). From west to east, these aquifers are the Edwards-Trinity (Plateau) Aquifer underlying the Edwards Plateau and Trans-Pecos regions, the Trinity Aquifer of the Hill Country region, and the Edwards Aquifer of the Balcones Fault Zone (BFZ). The BFZ is a Miocene-age series of predominantly southeast-dipping normal faults trending southwest to northeast along the eastern edge of the plateau (78, 79) (Fig. 1A). The resulting displacement along the largest faults formed the Balcones Escarpment, a prominent topographic feature that affects regional weather and stream drainage patterns along the southern and eastern margins of the Edwards Plateau (80, 81). The Edwards–Trinity system is bounded along its southeastern edge by a well-defined freshwater-saline water transition that minimizes flow of freshwater from the Edwards Aquifer (82) (Fig. 1A).

The Edwards Aquifer is the most hydrologically active aquifer in the Edwards-Trinity system (36), discharging about 1.1 km³ of water annually (83). The two southern segments of the Edwards (the Barton Springs and San Antonio segments) supply water to over 2.3 million people in the city of San Antonio and surrounding counties in south-central Texas and have been designated as sole-source aquifers for drinking water by the US Environmental Protection Agency (84). Aquifer thickness ranges from 60 to 180 m, with an average saturated thickness of 170 m in the southern portion (46). Regionally, the Edwards is subdivided by natural barriers into three segments. The San Antonio segment is separated from the Barton Springs segment by a groundwater divide near the city of Kyle in Hays County, and the Barton Springs segment is separated from the northern segment by the Colorado River (Fig. 1A). Locally, the southern segments of the Edwards Aquifer can be classified into three functionally distinct areas: the contributing zone, the recharge zone (Edwards outcrop), and the artesian or confined zone (Edwards subcrop). The contributing zone is the catchment area of the Edwards Plateau where precipitation infiltrates exposed Edwards and Trinity group rocks, migrating downward through the rock matrix by way of fractures and karst features such as sinkholes and directly recharging the water table (85) (Fig. 1B). Springs and seeps occur along the northern, eastern, and southern margins of the Edwards-Trinity (Plateau) Aquifer near the bases of the Edwards and Trinity groups where the water table intersects the land surface, discharging groundwater naturally to the surface under the force of gravity and providing base flow to streams that ultimately recharge the Edwards Aquifer downgradient (82, 86). Streams lose flow as they move across the exposed (unconfined) portion of the aquifer and recharge the water table through faults, fractures, and karst features. Unlike the southern segments of the Edwards, the northern segment lacks a distinct contributing zone; recharge occurs primarily through sinkholes on the aquifer outcrop (50, 87). In the confined zone, rock formations younger than Edwards cap the underlying aquifer, creating hydraulic pressure that forces water to the surface through the confining layer, creating natural discharge through artesian springs (Fig. 1*B*).

The unique geologic structure of the Edwards Aquifer controls the direction of groundwater movement, with permeable rock units horizontally adjacent to less-permeable units along fractures and faults that may act as either barriers or conduits to groundwater flow depending on the nature of the fault (88–90). Patterns of subsurface flow may be locally complex (91, 92), but in the southern segments water generally flows from areas of higher elevation in the southwest to discharge features in the northeast (36). Although the topographic boundaries between adjacent surface watersheds in the contributing zone of the Edwards Aquifer are well-defined, surface watershed boundaries may or may not be coincident with the extent of the underlying groundwater basins, especially where local geology is complex, for example in the northern segment of the Edwards (81, 93).

Results

Geographic Patterns of Population Structure. Population structure of Eurycea in the Edwards-Trinity system is consistent with a hierarchical island model (94, 95), in which local subpopulations are clustered into neighborhoods within watersheds united by gene flow. Hierarchical F statistics calculated for a two-level hierarchy (subpopulation within watershed, and watershed within subbasin) revealed significant effects on overall differentiation at one or both levels (SI Appendix, Tables S1–S3). Bayesian assignment (96) revealed well-defined population structure, with individuals always being strongly assigned to a given population. Below, we detail geographic patterns of population structure by major geographic regions of the Edwards-Trinity system (excluding the subgenus Typhlomolge) as follows: (i) northern segment of the Edwards (BFZ) Aquifer ("northern region"), (ii) southeastern Edwards (BFZ) Aquifer and its contributing zone in the Hill Country portion of the Trinity Aquifer ("southeastern"), and (iii) southwestern Edwards (BFZ) Aquifer and its contributing zone in the Edwards-Trinity (Plateau) Aquifer ("southwestern").

Northern region. Populations in the northern region have been classified as one of three distinct species (Eurycea chisholmensis, Eurycea naufragia, and Eurycea tonkawae) (60) that together comprise the highly divergent subgenus Septentriomolge (62). Hierarchical F statistics show a strong effect of subpopulation (within watershed) and watershed (within subbasin) on overall genetic structure (SI Appendix, Table S1). We analyzed population structure in a hierarchical fashion, beginning with all individuals and iteratively removing distinct clusters of individuals in subsequent analyses where we suspected additional subdivision exists. At the highest level (i.e., including all samples), individuals are strongly assigned to one of two population clusters (K) separated by the South Fork of the San Gabriel River (Fig. 2 and SI Appendix, Fig. S1). The northern cluster is further subdivided into three groups of populations, roughly corresponding to (from north to south) the Salado Creek, Berry Creek, and San Gabriel River (North and Middle forks) watersheds (Fig. 2 and SI Appendix, Fig. S2). The Salado Creek cluster includes the type locality for E. chisholmensis (Salado Springs) (60), whereas the North Fork of the San Gabriel River cluster includes the type locality for E. naufragia (Buford Hollow Spring) (60). Populations in the Berry Creek group have been assumed to represent E. naufragia based on geographic proximity to populations of that species, although most of these samples were collected subsequent to the most recent taxonomic revision in 2000 and had not been genotyped before this study. The southern cluster consists of populations classified as E. tonkawae, including the type locality at Stillhouse Hollow Springs (60) (Fig. 2). Within this cluster, indi-

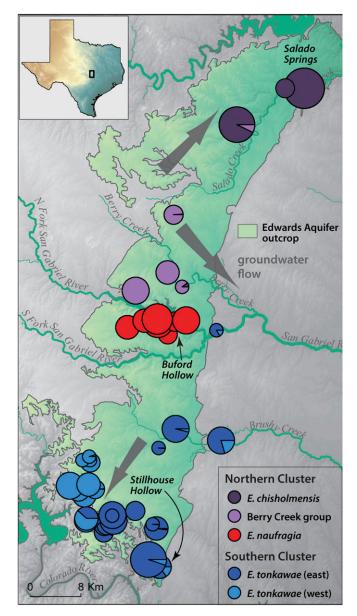


Fig. 2. Summary of hierarchical Structure analyses for the northern clade (subgenus Septentriomolge) showing the average individual membership coefficient (Q) for each sampling site. At the highest level of population structure (Pr(X|K=2)), individuals were assigned to northern and southern clusters separated by a hydrologic divide near the South Fork of the San Gabriel River. Each cluster is further subdivided, for a total of five major clusters shown here. Type localities for named species are shown. The size of each pie diagram is proportional to the number of sampled individuals per site (n = 1-9). See SI Appendix, Figs. S1-S3 and S12A and Tables S6 and S7 for further detail.

viduals are strongly assigned to one of two distinct population groups (Fig. 2 and SI Appendix, Fig. S3). The eastern group is found primarily in the Bull Creek watershed, with peripheral subpopulations in the Brushy, Shoal, and Walnut Creek watersheds. The narrowly distributed western group is restricted to the Buttercup Creek and Cypress Creek subwatersheds. These eastern and western groups come into narrow contact along the divide separating the Cypress Creek and Bull Creek drainages, with individuals from two populations (SAS Canyon and Kretschmarr Cave; SI Appendix, Fig. S12A) showing evidence of admixture. Southeastern region. Within the southeastern region, seven clus-

ters were inferred spanning (from north to south) the Middle

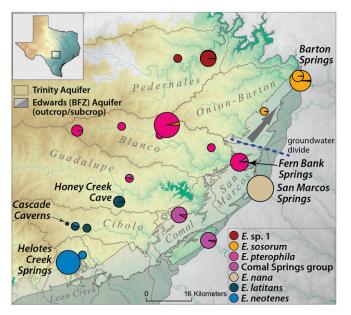


Fig. 3. Results of Structure analysis for the southeastern region (excluding the subgenus *Typhlomolge*) showing the average individual membership coefficient (Q) for each sampling site for a representative run at K=7. Type localities for named species are shown. Cascade Caverns (the type locality for *E. latitans*) was not sampled. The size of each pie diagram is proportional to the number of sampled individuals per site (n=1-11). See *SI Appendix*, Figs. S4 and S12B and Tables S6 and S7 for further detail.

Colorado, Guadalupe, and San Antonio river basins (Fig. 3). Some of these clusters correspond to named species, while others represent intraspecific population groupings (SI Appendix, Fig. S4). Within the Middle Colorado, two clusters were inferred, one in the Pedernales subbasin corresponding to an undescribed species (hereafter, E. sp. 1) and one in the Barton Springs segment of the Edwards Aquifer (the Barton and Onion Creek watersheds), corresponding to Eurycea sosorum (60). Within the Guadalupe basin, Eurycea nana—the San Marcos Springs endemic—forms a distinct cluster. Remaining populations in the Guadalupe and San Antonio basins belong to the Eurycea neotenes species complex, a group of four named taxa (Eurycea pterophila, E. neotenes, Eurycea latitans, and Eurycea tridentifera) that exhibit extensive incomplete lineage sorting and/or introgressive hybridization based on mtDNA sequence data (65). These populations form five clusters that are consistent with named species or population groups found mostly in separate watersheds: Blanco River (E. pterophila), Comal River (E. sp.), Leon Creek (E. neotenes), and the Cibolo Creek (E. latitans) plus Honey Creek (E. tridentifera) catchments. Individuals from Honey Creek Cave (the type locality for E. tridentifera) are strongly assigned to a cluster with individuals from two populations in the Cibolo Creek Headwaters watershed. Although we did not sample from the type locality of E. latitans (Cascade Caverns), the single sample from nearby Pfeiffer's Water Cave shows evidence of admixture between the E. latitans cluster and the Leon Creek E. neotenes cluster (SI Appendix, Fig. S4).

Southwestern region. Populations in the southwestern region belong to the Eurycea troglodytes species complex, a group of species found in the western Edwards and Edwards-Trinity (Plateau) aquifers (60). Sampled populations are strongly subdivided hierarchically by nested watershed (SI Appendix, Table S3). Three major, divergent clusters were inferred that correspond to distinct species (discussed below; see Fig. 4 and SI Appendix, Figs. S5 and S12C), and there is further subdivision within each species (SI Appendix, Figs. S6, S8, and S9). The easternmost cluster (hereafter, E. sp. 2) is found in the headwaters of the Guadalupe, upper North Medina, and Pedernales

River watersheds (Fig. 4 and *SI Appendix*, Fig. S12C). A second major cluster corresponding to *E. troglodytes* comprises populations found east of the Nueces River, in the eastern Nueces Headwaters, Frio, Hondo, and Upper West Medina watersheds (Fig. 4 and *SI Appendix*, Figs. S8 and S12C). The third, westernmost cluster (*E.* sp. 3) is found west of the Nueces River, in the Nueces Headwaters, West Nueces, Elm-Sycamore, and Lower Devils subbasins (Fig. 4 and *SI Appendix*, Figs. S9 and S12C).

Population and Species Tree Estimation. Maximum likelihood analysis (SI Appendix, Methods) and quartet inference under the multispecies coalescent model (97) of our largest assembly (74,955 SNPs) indicated similar relationships among populations; these two analyses (Fig. 5 and SI Appendix, Fig. S10) differ primarily in relationships within the southeastern region. These population trees are largely congruent with previous phylogenetic analyses of the group (60) but provide expanded coverage of the diversity and greater resolution of population assignment to species. The deepest division in the phylogeny separates northern and southern clades across the Colorado River and Mt. Bonnell fault of the BFZ. The northern clade (subgenus Septentriomolge) is represented by three named species (E. chisholmensis, E. naufragia, and E. tonkawae), two of which show clear evidence of additional geographic structuring across watersheds. Populations in the Berry Creek watershed are supported as the sister group of populations in the Salado Creek watershed (including topotypic E. chisholmensis samples) with strong support. Two geographic subgroups are also apparent within E. tonkawae, corresponding to eastern and western clusters of populations, but this division is not strongly supported. South of the Colorado River, the next deepest divergence separates the subterranean subgenus Typhlomolge (represented in our analysis by Eurycea waterlooensis and Eurycea rathbuni) from the remaining species. The remaining species (subgenus Blepsimolge) are split into strongly supported southeastern and southwestern clades. Population groupings within these clades are consistent with clusters recovered in Structure analyses. Within the southwestern clade, three major lineages were inferred with strong support. Within the southeastern clade, six major lineages were inferred, although relationships among

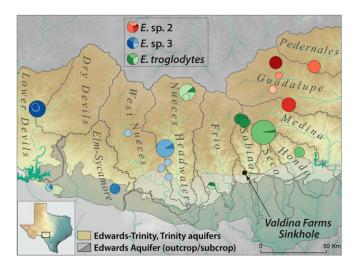


Fig. 4. Summary of hierarchical Structure analyses for the southwestern region showing the average individual membership coefficient (Q) for each sampling site. The type locality for *E. troglodytes* (Valdina Farms Sinkhole) is shown but was not sampled. Three major clusters were inferred corresponding to distinct species, each of which is further subdivided. The size of each pie diagram is proportional to the number of sampled individuals per site (n = 1-11). See *SI Appendix*, Figs. S5–S9 and S12C and Tables S6 and S7 for further detail.



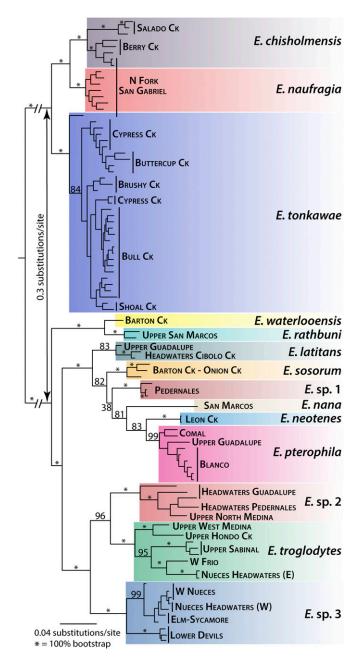


Fig. 5. Maximum likelihood tree of populations inferred from 74,955 SNPs. Branch labels are bootstrap support values from 1,000 replicates. See *SI Appendix*, Fig. S11 for the same topology with subpopulation names shown. Watershed boundaries (*SI Appendix*, Table S6) are from the National Hydrography Dataset Plus (https://www.epa.gov/waterdata/get-data).

these lineages are not well-resolved, especially within the *E. neotenes* species complex.

Species Delimitation Under Bayesian Phylogenetics and Phylogeography. Consistent with other studies that have used Bayesian Phylogenetics and Phylogeography (BPP) analysis (98, 99), the prior for θ had considerable impact on species delimitation results (*SI Appendix*, Table S4). Across analyses under different priors, the ranking of the models usually remained the same, although the posterior probabilities for alternative delimitations varied. For the northern clade, we mapped individuals to five population groups (based on Structure results; see Fig. 2) representing potential species for BPP analysis: Salado Creek, Berry Creek, San Gabriel River, eastern E.

tonkawae, and western E. tonkawae. Across prior specifications, the posterior probability that these populations represent distinct species ranged from 0.71 to 1 (SI Appendix, Table S4). Berry Creek, Salado Creek, and San Gabriel River populations were usually recovered as distinct species with probability 1, whereas eastern and western E. tonkawae populations were usually grouped together. In the southeastern clade, individuals were mapped to nine population groups: Comal Springs, Cibolo Creek, San Marcos Springs, Leon Creek, Lower Blanco, Upper Blanco, Barton Springs segment, Pedernales River, and Honey Creek. Different specifications of the priors resulted in support for six to nine species. Blanco River populations were recovered as belonging to the same species, usually along with the Comal Springs and nearby Bear Creek Spring populations. Under a six-species model, the Cibolo Creek and Honey Creek population groups were grouped together as belonging to the same species. In the southwestern clade, all three population groups mapped as potential species (Fig. 4) were inferred to represent distinct species with probability 1 (SI Appendix, Table S4).

Discussion

Hydrogeology Drives Population Divergence in the Edwards-Trinity **Eurycea.** Patterns of population divergence and speciation in the west-central Texas Eurycea have been driven in part by the complex surface and subsurface hydrogeology of the dynamic Edwards–Trinity system (64). At a regional scale, the boundaries of major clades coincide with major aquifer borders and distinct aquifer segments separated by hydrologic divides and the expression of major faults in the BFZ. Separation between the northern and southern clades across the Colorado River and Mt. Bonnell fault reflects deep evolutionary divergence, dating to at least the Middle Miocene (60). Population structure follows a hierarchical island model (95), where local subpopulations within nested catchment areas are united by gene flow. Patterns of species endemism range from single-site endemics to lineages that have widespread ranges spanning two or more subbasins. Species boundaries are frequently coincident with surface or subsurface hydrologic divides, consistent with a model of vicariant allopatric divergence and speciation. Alongside this nonadaptive population divergence are multiple instances of adaptive divergence between surface and subsurface intraspecific populations resulting in phenotypic convergence that has confounded previous species delimitation efforts based on morphology alone (69).

The intense faulting and fracturing in the BFZ and resulting structural controls on regional groundwater movement have driven population subdivision, and ultimately species formation, in this clade. In the northern region, subsurface groundwater divides separate distinct genetic clusters. The northernmost divide occurs between the Berry Creek and Salado Creek watersheds. North of this groundwater divide, groundwater follows northeast-trending faults and flows to the northeast, whereas south of this divide, groundwater in the Berry Creek drainage generally flows east (87) (Fig. 2). A second groundwater divide between Brushy Creek and the South Fork of the San Gabriel River similarly divides E. naufragia from E. tonkawae populations to the south on the Jollyville Plateau. South of this divide, groundwater flows southward, where it is captured by the deeply incised Colorado River along the southern margin of the Edwards Plateau (100). Faulting and an associated fault-bound relay ramp structure appear responsible for the east–west genetic subdivision within E. tonkawae, a pattern that is shared with other karst endemic species in the region (101). Cave populations in the Buttercup Creek karst (102) exhibit distinct troglomorphy and were hypothesized to represent a distinct species; these populations were only provisionally assigned to E. tonkawae (60). However, individuals from surface-dwelling populations in the Cypress Creek watershed are also strongly assigned to this same cluster, suggesting troglomorphic Buttercup Creek cave populations represent another instance of convergence (69). Dye tracing results have revealed subsurface connections between caves in the Buttercup Creek karst and

springs in the Cypress Creek watershed (103, 104) that may allow for gene flow between these separate drainages.

Local aquifer hydrodynamics are complex in the southeastern portion of the study area, where groundwater flow and recharge to the Edwards Aquifer is complicated by the structure of faults and the stratigraphy of rocks (105, 106). Water recharging the aquifer moves toward the northeast, along and parallel to northeast-trending faults, discharging at Comal, San Marcos, and Barton Springs. Surface and subsurface hydrologic divides serve as geographic barriers, but some are transient in nature. For example, a dynamic groundwater divide along Onion Creek near Kyle (Fig. 3) separates E. sosorum at the southern portion of its range from parapatric populations of E. pterophila in the Blanco River subbasin. North of this divide, groundwater in the Onion Creek watershed recharging the Edwards Aquifer usually flows northeast toward Barton Springs under average flow conditions (92). However, dye tracing has shown that during wet conditions groundwater can reverse direction, flowing up a structural dip toward San Marcos Springs instead of Barton Springs (92). This shifting groundwater divide may result in rare gene flow between watersheds, which may explain the pattern of mitochondrial DNA similarity between some individuals of E. sosorum from Barton Springs and E. nana at San Marcos Springs (65).

Taxonomic Implications. Our results have significant implications for the taxonomy of Edwards–Trinity Eurycea species, including ESA-listed and ESA-candidate species (*SI Appendix*, Table S5). In determining what constitutes a species, we view the various criteria used in delimiting species as contingent properties of metapopulation lineages that are evolving independently from other such lineages (107). Nonetheless, these contingent properties serve as important lines of evidence for assessing the evolutionary independence of lineages, including assessing their potential for gene flow or continued divergence. In determining which metapopulation lineages are evolving independently, it is useful to consider the evolutionary processes that result in lineage divergence, that is, natural selection, mutation, gene flow, and genetic drift (107). Of these evolutionary processes, we can say the most about gene flow based on available genetic data, although in some cases determining whether shared genetic variation is due to recent gene exchange or simply retained ancestral polymorphism following separation is challenging, particularly when populations have diverged only recently (108).

In the northern study area, our phylogenetic results show that all populations north of the San Gabriel River in and near the Berry Creek watershed are more closely related to *E. chisholmensis* than to *E. naufragia*, and so we assign these populations to *E. chisholmensis* (Fig. 6). These populations were initially assigned to *E. naufragia* (based on geographic proximity) when the three northern species were listed under the ESA in 2013 (56). Although the Berry Creek populations could be recognized as a new distinct species, rather than treated as a population segment of *E. chisholmensis*, there is evidence of gene flow or shared ancestral polymorphism between the Berry Creek and Salado Creek population groups (Fig. 2). Therefore, we consider these two population groups to be conspecific.

Our results also support assignment of the single salamander that is available from Georgetown Springs to *E. tonkawae* (*SI Appendix*, Figs. S1 and S11). However, this individual shows some indication of shared alleles with *E. naufragia* (*SI Appendix*, Fig. S1), suggesting possible past gene flow between *E. naufragia* and *E. tonkawae*. Nonetheless, this individual is not the result of a recent hybridization event, as it is genetically more similar to other individuals of *E. tonkawae* than any individuals of *E. naufragia* (*SI Appendix*, Fig. S11). This population may now be extinct, as we have been unable to locate any additional salamanders at Georgetown Springs since 1991.

Reassigning the Berry Creek and Georgetown Springs populations to other species significantly changes the known distribution of *E. naufragia* and necessitates revision of critical habitat designations under the ESA for *E. chisholmensis*, *E. naufragia*,

and *E. tonkawae*. Our results restrict the range of *E. naufragia* to springs south and east of Lake Georgetown in the North and Middle forks of the San Gabriel River watershed; all known populations of *Eurycea* north of Lake Georgetown are here included within *E. chisholmensis* (Fig. 6).

In the southeastern study area, our results suggest that specieslevel diversity has been overestimated within the E. neotenes species complex. Species in this group have been distinguished primarily by allozyme allele frequency differences (60). The population from Comal Springs has been suggested to represent a distinct species (60), and a petition for its listing has been filed under the ESA (109). However, individuals from Comal Springs and nearby Bear Creek Spring are genetically similar and share genetic variation with populations in the lower Blanco River and Guadalupe subbasins assigned to *E. pterophila* (see also ref. 110). We therefore recommend including both the Comal Springs and Bear Creek Spring populations within E. pterophila rather than recognizing them as a distinct, undescribed species (Fig. 6). Our results also show that E. tridentifera populations from the type locality at Honey Creek Cave group with E. latitans individuals from very near the type locality for that species at Cascade Caverns (Fig. 3), with little evidence to suggest that these two species are distinct. We therefore recommend that the name E. tridentifera (67) be subsumed under the older name E. latitans (111). Admixture between E. latitans and Leon Creek E. neotenes populations at Pfeiffer's Water Cave warrants further investigation with increased sampling (see also ref. 112).

In the southwestern study area, we recognize three species, two of them new. We restrict the name *E. troglodytes* to populations found east of the Nueces River, primarily in the Nueces Headwaters, Upper Frio, Hondo, and Medina subbasins (Fig. 6 and *SI Appendix*, Table S6). *Eurycea* sp. 2 appears to be restricted to springs in the headwaters of the Guadalupe, Pedernales, and North Prong of the Medina rivers. *Eurycea* sp. 3 is found west of the Nueces River, in the Nueces Headwaters, West Nueces, Elm-Sycamore, and Lower Devils subbasins (Fig. 6 and *SI Appendix*, Table S6).

It is important to note that, although the species rank is the usual unit of currency for decision makers tasked with conservation prioritization, a focus on the species level alone ignores geographic variation and population divergence within species such as *E. chisholmensis*, *E. tonkawae*, *E. pterophila*, *E. troglodytes*, and *E.* sp. 3 (Figs. 2–4). Genetically divergent and geographically separated populations within these species (some of which have previously been considered to be full species) may warrant recognition as "distinct population segments" as defined under the ESA.

Implications for Groundwater and Biodiversity Conservation. The region bordering the eastern half of the Edwards Aquifer from San Antonio to the greater Austin area includes some of the fastest-growing counties and metropolitan areas in the United States (47, 113). Although the city of Austin does not rely directly on groundwater to meet its water demands, the city of San Antonio and many smaller municipalities rely largely or exclusively on groundwater from the Edwards Aquifer (114). Significant water table drawdown has already occurred over large portions of the study area, especially in the northern segment of the Edwards Aquifer (114, 115). From 1980 to 2000, groundwater pumping for municipal, rural domestic, and industrial use in the northern segment of the Edwards Aquifer nearly doubled (114). Between 1966 and 1990, over 50 m of drawdown occurred in some areas (116).

Major drawdown has also occurred over portions of the Trinity Aquifer. Groundwater yields from the Trinity Aquifer are about 250 times less than average yields in the adjacent Edwards Aquifer (117). Intensive pumping in the Trinity over the last several decades has resulted in water table declines, decreased well yields, and diminished baseflow to springs and streams (48, 118). The US Geological Survey, in cooperation with the US Fish and Wildlife Service, investigated the potential impacts of

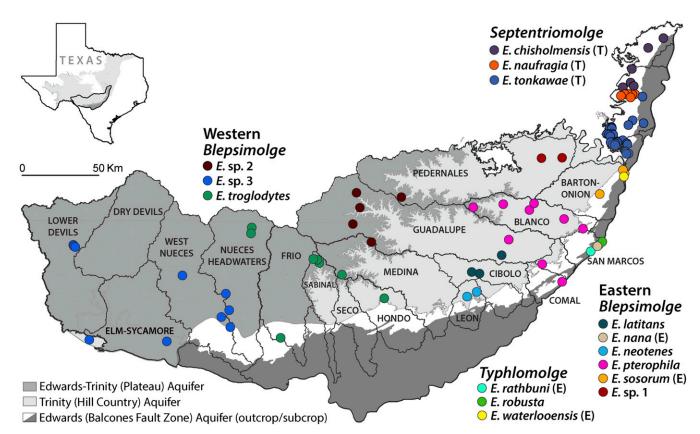


Fig. 6. Map of sampled populations and species assignments inferred from results presented here, along with major watersheds in the study area. Species currently listed as Threatened (T) or Endangered (E) under the ESA are indicated.

groundwater pumping on known Eurycea spring localities located in the middle zone of the Trinity Aquifer (hereafter, Middle Trinity) (119). The Middle Trinity zone was chosen because it was the focus of a numerical groundwater availability model for the Trinity Aquifer (117). For Middle Trinity springs, projected water levels for both average recharge conditions and droughtof-record (DOR) conditions show drawdown at every spring. Under average recharge conditions projected for 2050, water table levels at 15 of 19 springs are predicted to decline by more than 3 m, whereas under DOR conditions water levels at 12 of 19 springs are projected to decline by more than 15 m (119).

Groundwater depletion, which results in diminished springflow and water table declines, is expected to have significant negative impacts on biodiversity and ecosystem functioning (30, 120, 121). This is especially true in shallow, dynamic karst aquifers (122) like those in the Edwards-Trinity system. For groundwater-obligate species such as Eurycea salamanders, declining water tables and reduced springflow causes habitat loss at the surface and subsurface, resulting in declines in individual abundance and eventually population extinction. Extirpations and functional extinctions (123) may then reverberate through ecological networks via complex interactions among species (see, e.g., refs. 31 and 32). In the worst case, population losses may result in the impaired functioning of groundwaterdependent ecosystems (see, e.g., refs. 33–35).

Effective conservation of groundwater-dependent biodiversity and ecosystem services faces a number of major challenges. The unique hydrogeologic properties of karstic aquifers that make them a valuable water source also make them especially vulnerable to contamination (124, 125). In west-central Texas, rapid population growth, increased water demands (126), and a warming climate with more frequent drought (127) are placing increased stress on limited water resources for human needs. During droughts, enforcement of the ESA requires that minimum environmental flows be maintained to prevent the unlawful "take" of the Edwards Aquifer's endangered species (128). However, state law in Texas treats surface water and groundwater as separate resources (despite their functional interdependence), with groundwater considered private property (129). Under this so-called rule of capture law, there is no enforceable legal mandate at the state or local level to maintain minimum aquifer levels (and hence springflow and stream baseflow) needed by endangered species (130). Without joint management of surface and subsurface waters as a single common-pool resource, the aquifers, springs, and streams of the Edwards-Trinity and the regional ecosystems they sustain will become increasingly threatened.

Conclusions

Global change driven by human population growth (131), agricultural expansion (132), and climate change (133, 134) is steadily increasing the demand for freshwater, compounding the severe water scarcity already faced by an estimated 4 billion people (135). Groundwater is critically important for human health and ecosystem functioning in many semiarid regions because of low rainfall, frequent droughts, and little permanent surface water. The conservation of groundwater biodiversity should be a low-cost by-product of water conservation programs (29), but groundwater depletion increasingly threatens biodiversity as well as the stability of aquatic, terrestrial, and subsurface ecosystems. Because most groundwater species are difficult to monitor, abundance declines are likely to go unnoticed until after populations have already been lost. Regional climate models that predict increased air temperature (136, 137), together with hydrologic models that project decreased springflow (119, 138, 139), predict that Edwards-Trinity Eurycea salamander populations and other codistributed groundwater-dependent species are highly vulnerable to extinction within the next century.

ny occupied on December 30, 2021

Methods

Sampling. We sampled 1–11 individuals from 99 spring or cave sites for a total of 303 samples representing all known species of Edwards–Trinity Eurycea, except E. (Typhlomolge) robusta (which is known from a single specimen collected in the 1950s) (SI Appendix, Fig. S12 and Table S6). Most specimens and tissues were collected previously and some have been analyzed for allozyme and/or mtDNA variation (60, 62, 65). Specimens, collecting data, and remaining tissue samples are housed in the Genetic Diversity Collection of the Biodiversity Center, Department of Integrative Biology, The University of Texas at Austin. Tissues and specimens were collected following The University of Texas at Austin IACUC protocol AUP-2018-00151.

Restriction-Site-Associated DNA Sequencing. Sequence data were collected using double-digest restriction-site-associated DNA sequencing (ddRADseq) following the protocol of Peterson et al. (140). Libraries were constructed and sequenced at the Genomic Sequencing and Analysis Facility, The University of Texas at Austin, on the Illumina HiSEq. 2500 platform (100-bp paired-end run). Additional details are provided in *SI Appendix, Methods*.

RAD Data Assembly. We used both the Stacks computational pipeline (141) and ipyrad (142) to filter and sort reads, identify loci de novo, and genotype individuals. Stacks was used to produce SNP-based datasets for phylogenetic and population structure inference using first reads only because they were of higher quality. Both first and second reads were used to identify full sequence RAD loci with ipyrad for BPP analyses. Because the parameters used in de novo locus identification and genotyping may affect downstream analyses and resulting inference (143–145), we examined a range of values for parameters to optimize assembly in both pipelines. Additional details are provided in *SI Appendix, Methods*. RAD data assembly was performed on the Lonestar 5 high performance computing system at the Texas Advanced Computing Center, The University of Texas at Austin. Demultiplexed fastq files are available in the NCBI Sequence Read Archive (SRA) under study number PRINA504898 (146). Raw data, barcodes, and assemblies are available in the Dryad data package associated with this article (147).

Population Structure Analysis. We assigned individuals to species based on multiple lines of evidence, including morphology, allozymes, mtDNA, and geographic distribution (60, 62, 65). For individuals from populations whose taxonomic assignment was uncertain, we used the Bayesian clustering method implemented in Structure (96) to assign individuals to species based on their multilocus genotype. Additional details are provided in *SI Appendix, Methods.* We also estimated hierarchical *F* statistics (subpopulation within watershed within subbasin) following ref. 148 and tested the statistical significance of these levels on population differentiation using a generalized likelihood-ratio test (149) implemented in the HierFstat package (150, 151) for R (152).

- United Nations (2003) Water for people water for life (United Nations Educational, Scientific and Cultural Organization, Paris).
- Siebert S, et al. (2010) Groundwater use for irrigation—A global inventory. Hydrol Earth Syst Sci 14:1863–1880.
- 3. Danielopol DL, Pospisil P, Rouch R (2000) Biodiversity in groundwater: A large-scale view. *Trends Ecol Evol* 15:223–224.
- Gibert J, Deharveng L (2002) Subterranean ecosystems: A truncated functional biodiversity. Bioscience 52:473–481.
- 5. Griebler C, Avramov M (2015) Groundwater ecosystem services: A review. *Freshw Sci*
- 34:355–367.

 6. Foster SSD, Chilton PJ (2003) Groundwater: The processes and global significance of

aquifer degradation. Philos Trans R Soc Lond B Biol Sci 358:1957-1972.

- Giordano M (2009) Global groundwater? Issues and solutions. Annu Rev Environ Resour 34:153–178.
- Van der Gun J (2012) Groundwater and global change: Trends, opportunities, and challenges (United Nations World Water Assessment Programme, Paris),pp 1–44.
- Aeschbach-Hertig W, Gleeson T (2012) Regional strategies for the accelerating global problem of groundwater depletion. Nat Geosci 5:853–861.
- Gleeson T, Wada Y, Bierkens MF, van Beek LP (2012) Water balance of global aquifers revealed by groundwater footprint. Nature 488:197–200.
- 11. Famiglietti JS (2014) The global groundwater crisis. *Nat Clim Chang* 4:945–948.
- Fan Y, Li H, Miguez-Macho G (2013) Global patterns of groundwater table depth. Science 339:940–943.
- Konikow LF, Kendy E (2005) Groundwater depletion: A global problem. Hydrogeol J 13:317–320.
- Dalin C, Wada Y, Kastner T, Puma MJ (2017) Groundwater depletion embedded in international food trade. *Nature* 543:700–704.
- Castle SL, et al. (2014) Groundwater depletion during drought threatens future water security of the Colorado River Basin. Geophys Res Lett 41:5904–5911.

Phylogenetic Inference. We used the SVDquartets inference method of Chifman and Kubatko (97) implemented in PAUP* v4 (153) to estimate the species tree and a population tree under the multispecies coalescent model using the 74,955-locus SNP dataset. This method assumes that any incongruence between an SNP genealogy and the species tree is due only to the stochastic coalescent process and uses the full data directly, incorporating both mutational and coalescent variance inherent in species-tree estimation (154). The observed site pattern distribution is used to infer the true split for quartets of taxa based on the best singular value decomposition (SVD) score (97), followed by the quartet assembly algorithm of Reaz et al. (155) for tree inference. We evaluated 100,000 random quartets, both with and without the Erik+2 normalization method of Fernández-Sánchez and Casanellas (156). Ambiguous changes were treated as missing. Nonparametric bootstrapping with 10,000 replicates was used to quantify uncertainty in the inferred splits and a 50% majority-rule consensus tree was constructed in PAUP* and visualized in FigTree (tree.bio.ed.ac.uk/software/figtree/).

We also inferred the maximum likelihood phylogeny of the 74,955-locus dataset using RAxML v.8 (157) with a conditional likelihood method (158) that corrects for the exclusion of invariant sites (159) (-m ASC_GTRGAMMA–asc-corr lewis). To find the best ML tree, the matrix was analyzed 1,000 times and the best tree was kept; 1,000 bootstrap replicates were performed to assess support, and we used the SumTrees program in the DendroPy Python library (160) to summarize bootstrap support values. RAxML analyses were performed on the CIPRES Science Gateway server v3.2 (161).

BPP Species Delimitation. We used the Bayesian Markov chain Monte Carlo method of Yang and Rannala implemented in the program BPP v3.3a (162–165) to generate the posterior probabilities of different species delimitations based on support for recent coalescence. This method is a full likelihood-based implementation of the multispecies coalescent model, accounting for the species phylogeny while accommodating uncertainty due to unknown gene trees (topologies and branch lengths) at individual loci, and incomplete lineage sorting. Assumptions include no recombination within a locus and free recombination between loci, neutral evolution at a constant rate at each locus under the JC69 model of nucleotide substitution, and no gene flow between species (165). The data for these analyses are full sequences (including invariant sites) that are 108 bp in length. Additional details are provided in *SI Appendix, Methods*.

ACKNOWLEDGMENTS. We thank Craig Farquhar of the Texas Parks and Wildlife Department for assistance; Nathan Bendik, Paul Chippindale, Andy Gluesenkamp, John Karges, Lisa O'Donnell, Chad Norris, Colin Peden, and the late Andy Price for tissue samples; Deren Eaton and Isaac Overcast for assistance and advice with their program ipyrad; and Joe Allen, Richard Evans, Joe Garcia, Doug James, Antia Lamas-Linares, Jawon Song, and Virginia Truehart for assistance with Texas Advanced Computing Center resources. This work was funded by the US Fish and Wildlife Service, Section 6 Grant TX E158-R-1, through Texas Parks and Wildlife Department Contract 443022.

- Scanlon BR, et al. (2012) Groundwater depletion and sustainability of irrigation in the US High Plains and Central Valley. Proc Natl Acad Sci USA 109:9320–9325.
- Garrick DE, et al. (2017) Valuing water for sustainable development. Science 358: 1003–1005.
- Fenichel EP, et al. (2016) Measuring the value of groundwater and other forms of natural capital. Proc Natl Acad Sci USA 113:2382–2387.
- Herman JS, Culver DC, Salzman J (2001) Groundwater ecosystems and the service of water purification. Stanf Environ Law J 20:479–495.
- Meckenstock RU, et al. (2015) Biodegradation: Updating the concepts of control for microbial cleanup in contaminated aquifers. Environ Sci Technol 49:7073–7081.
- 21. Alexander M (1981) Biodegradation of chemicals of environmental concern. *Science*
- 22. Engel AS, et al. (2010) Linking phylogenetic and functional diversity to nutrient spiraling in microbial mats from Lower Kane Cave (USA). ISME J 4:98–110.
- Hutchins BT, Engel AS, Nowlin WH, Schwartz BF (2016) Chemolithoautotrophy supports macroinvertebrate food webs and affects diversity and stability in groundwater communities. *Ecology* 97:1530–1542.
- Fiser C, Pipan T, Culver DC (2014) The vertical extent of groundwater metazoans: An
 ecological and evolutionary perspective. Bioscience 64:971–979.
- Delić T, Trontelj P, Rendoš M, Fišer C (2017) The importance of naming cryptic species and the conservation of endemic subterranean amphipods. Sci Rep 7:3391.
- Gallagher AJ, Hammerschlag N, Cooke SJ, Costa DP, Irschick DJ (2015) Evolutionary theory as a tool for predicting extinction risk. *Trends Ecol Evol* 30:61–65.
 Kotiaho JS, Kaitala V, Komonen A, Päivinen J (2005) Predicting the risk of extinction
- from shared ecological characteristics. *Proc Natl Acad Sci USA* 102:1963–1967.

 28. McKinney ML (1997) Extinction vulnerability and selectivity: Combining ecological
- and paleontological views. Annu Rev Ecol Syst 28:495–516.
- Sket B (1999) High biodiversity in hypogean waters and its endangerment: The situation in Slovenia, the dinaric karst, and Europe. Crustaceana 72:767–779.

- Boulton AJ, Fenwick GD, Hancock PJ, Harvey MS (2008) Biodiversity, functional roles and ecosystem services of groundwater invertebrates. *Invertebr Syst* 22:103–116.
- 31. Johnson CN, et al. (2017) Biodiversity losses and conservation responses in the Anthropocene. Science 356:270–275.
- 32. Estes JA, et al. (2011) Trophic downgrading of planet Earth. Science 333:301-306.
- Duffy JE, Godwin CM, Cardinale BJ (2017) Biodiversity effects in the wild are common and as strong as key drivers of productivity. Nature 549:261–264.
- 34. Rockström J, et al. (2009) A safe operating space for humanity. Nature 461:472–475.
- Hautier Y, et al. (2015) Anthropogenic environmental changes affect ecosystem stability via biodiversity. Science 348:336–340.
- Kuniansky EL, Ardis AF (2004) Hydrogeology and ground-water flow in the Edwards-Trinity aquifer system, west-central Texas: Regional aquifer-system analysis— Edwards-Trinity. US Geological Survey Professional Paper 1421-C (US Geological Survey, Reston, VA), p 85.
- Bowles DE, Arsuffi TL (1993) Karst aquatic ecosystems of the Edwards Plateau region
 of central Texas, USA: A consideration of their importance, threats to their existence,
 and efforts for their conservation. Aquat Conserv 3:317–329.
- Longley G (1981) The Edwards Aquifer: Earth's most diverse groundwater ecosystem? Int J Speleol 11:123–128.
- Barr CB, Spangler PJ (1992) A new genus and species of stygobiontic dryopid beetle, Stygoparnus comalensis (Coleoptera: Dryopidae), from Comal Springs, Texas. Proc Biol Soc Wash 105:40–54.
- Spangler PJ, Barr CB (1995) A new genus and species of stygobiontic dytiscid beetle, Comaldessus stygius (Coleoptera: Dytiscidae: Bidessini) from Comal Springs, Texas. Insecta Mundi 9:301–308.
- Jean A, Telles ND, Gibson JR, Foley D, Miller KB (2012) Description of a new genus and species of stygobiontic diving beetle, *Psychopomporus felipi* Jean, Telles, and Miller (Coleoptera: Dytiscidae: Hydroporinae), from the Edwards-Trinity aquifer system of Texas, USA. *Coleopt Bull* 66:105–110.
- Hershler R, Longley G (1986) Phreatic hydrobiids (Gastropoda: Prosobranchia) from the Edwards (Balcones Fault Zone) Aquifer region, south-central Texas. Malacologia 27:127–172.
- 43. Hutchins BT (2017) The conservation status of Texas groundwater invertebrates. Biodivers Conserv 27:475–501.
- Mace RE, Angle ES (2004) Aquifers of the Edwards Plateau. Texas Water Development Board Report 360 (Texas Water Development Board, Austin, TX), pp 1–20.
- Anaya R (2004) Conceptual model for the Edwards-Trinity (Plateau) Aquifer system, Texas. Texas Water Development Board Report 360: Aquifers of the Edwards Plateau (Texas Water Development Board, Austin, TX), pp 22–62.
- George PG, Mace RE, Petrossian R (2011) Aquifers of Texas. Texas Water Development Board Report 380 (Texas Water Development Board, Austin, TX), pp 1–182.
- US Census Bureau (2016) Five of the nation's eleven fastest-growing cities are in Texas (US Census Bureau, Suitland, MD). Available at https://www.census.gov/ newsroom/press-releases/2016/cb16-81.html. Accessed December 18. 2018.
- Ashworth JB (1983) Ground-water availability of the lower Cretaceous formations in the Hill Country of south-central Texas. Texas Department of Water Resources Report 273 (Texas Department of Water Resources, Austin, TX), pp 1–39.
- Chowdhury AH (2009) GAM Run 08-70. GAM run report (Texas Water Development Board, Austin, TX), pp 1–42.
- Boghici R (2008) Changes in water levels in Texas, 1990–2000. Texas Water Development Board Report 371 (Texas Water Development Board, Austin, TX), pp 1–42.
- Brune G (1975) Major and historical springs of Texas. Texas Water Development Board Report 189 (Texas Water Development Board, Austin, TX), pp 1–91.
- RECON Environmental, Inc; Hicks & Company; Zara Environmental, LLC; BIO-WEST (2012)
 Edwards aquifer recovery implementation program habitat conservation plan, pp 1–414.
- National Research Council (2015) Review of the Edwards Aquifer Habitat Conservation Plan (National Academies Press, Washington, DC).
- National Academies of Sciences, Engineering, and Medicine (2017) Review of the Edwards Aquifer Habitat Conservation Plan: Report 2 (National Academies Press, Washington, DC).
- Bendik NF, Sissel BN, Fields JR (2014) Effect of urbanization on abundance of Jollyville Plateau salamanders (Eurycea tonkawae). Herpetol Conserv Biol 9:206–222.
- 56. US Fish and Wildlife Service (2014) Endangered and threatened wildlife and plants; determination of threatened species status for the Georgetown Salamander and Salado Salamander throughout their ranges; final rule. Fed Regist 79:10236–10293.
- US Fish and Wildlife Service (1997) Endangered and threatened wildlife and plants; final rule to list the Barton Springs Salamander as endangered. Fed Regist 62:23377–23392.
- 58. US Fish and Wildlife Service (1980) Endangered and threatened wildlife and plants; listing of the San Marcos Salamander as threatened, the San Marcos Gambusia as endangered, and the listing of critical habitat for Texas wild rice, San Marcos Salamander, San Marcos Gambusia, and fountain darter. Fed Regist 45:47355–47364.
- Hubbs C, Edwards RJ, Garrett GP (2008) An annotated checklist of the freshwater fishes of Texas, with keys to identification of species (Texas Academy of Science, Edinburg, TX), pp 1–44.
- Chippindale PT, Price AH, Wiens JJ, Hillis DM (2000) Phylogenetic relationships and systematic revision of central Texas hemidactyliine plethodontid salamanders. Herpetol Monogr 14:1–80.
- Chippindale PT, Price AH, Hillis DM (1993) A new species of perennibranchiate salamander (Eurycea: Plethodontidae) from Austin, Texas. Herpetologica 49:248–259.
- Hillis DM, Chamberlain DA, Wilcox TP, Chippindale PT (2001) A new species of subterranean blind salamander (Plethodontidae: Hemidactyliini: Eurycea: Typhlomolge) from Austin, Texas, and a systematic revision of central Texas paedomorphic salamanders. Herpetologica 57:266–280.
- Baker JK (1961) Distribution of and key to the neotenic Eurycea of Texas. Southwest Nat 6:27–32.

- Sweet SS (1978) The evolutionary development of the Texas Eurycea (Amphibia: Plethodontidae). PhD dissertation (Univ of California, Berkeley, CA).
- Bendik NF, Meik JM, Gluesenkamp AG, Roelke CE, Chippindale PT (2013) Biogeography, phylogeny, and morphological evolution of central Texas cave and spring salamanders. BMC Evol Biol 13:201.
- Potter FE, Jr, Sweet SS (1981) Generic boundaries in Texas cave salamanders, and a redescription of Typhlomolge robusta (Amphibia: Plethodontidae). Copeia 1981:64–75.
- Mitchell RW, Reddell JR (1965) Eurycea tridentifera, a new species of troglobitic salamander from Texas and a reclassification of Typhlomolge rathbuni. Tex J Sci 17:12–27.
- Mitchell RW, Smith RE (1972) Some aspects of the osteology and evolution of the neotenic spring and cave salamanders (Eurycea, Plethodontidae) of central Texas. Tex 1 Sci 23:343–362.
- Wiens JJ, Chippindale PT, Hillis DM (2003) When are phylogenetic analyses misled by convergence? A case study in Texas cave salamanders. Syst Biol 52:501–514.
- Bickford D, et al. (2007) Cryptic species as a window on diversity and conservation. Trends Ecol Evol 22:148–155.
- Salzman J (1990) Evolution and application of critical habitat under the Endangered Species Act. Harvard Environ Law Rev 14:311–342.
- 72. US Fish and Wildlife Service; National Marine Fisheries Service (2016) Listing endangered and threatened species and designating critical habitat; implementing changes to the regulations for designating critical habitat; final rule. Fed Regist 81:7414–7440.
- Daugherty CH, Cree A, Hay JM, Thompson MB (1990) Neglected taxonomy and continuing extinctions of tuatara (Sphenodon). Nature 347:177–179.
- O'brien SJ, Mayr E (1991) Bureaucratic mischief: Recognizing endangered species and subspecies. Science 251:1187–1188.
- Puig-Williams V (2015) The endangered springflow act: How the endangered species
 act influences groundwater law and protects springflow in Texas. SSRN Electron J,
 10.2139/ssrn.2540317.
- Puig-Williams V (2016) Regulating unregulated groundwater in Texas: How the state could conquer this final frontier. Tex Water J 7:85–96.
- Votteler TH (1998) The little fish that roared: The Endangered Species Act, state groundwater law, and private property rights collide over the Texas Edwards Aquifer. Environ Law 28:845–880.
- 78. Sellards EH, Baker CL (1935) The Geology of Texas. Volume II: Structural and Economic Geology (Univ of Texas at Austin, Austin, TX), Vol 2, p 884.
- Weeks AW (1945) Balcones, Luling, and Mexia fault zones in Texas. Am Assoc Pet Geol Bull 29:1733–1737.
- Caran SC, Baker VR (1986) Flooding along the Balcones escarpment, central Texas.
 The Balcones Escarpment, Central Texas, eds Abbott PL, Woodruff CM, Jr (Geological Society of America, Boulder, CO), pp 1–14.
- Woodruff CM, Jr, Abbott PL (1986) Stream piracy and evolution of the Edwards aquifer along the Balcones escarpment, central Texas. The Balcones Escarpment, Central Texas, ed Woodruff CM, Jr (Geological Society of America, Boulder, CO), pp 77–90.
- Barker RA, Bush PW, Baker ETJ (1994) Geologic history and hydrogeologic setting of the Edwards-Trinity aquifer system, west-central Texas. US Geological Survey Water Resources Investigations Report 94-4039 (US Geological Survey, Reston, VA), pp 1–55.
- 83. Smith BA, Hunt BB (2013) Enhanced recharge to the Barton springs segment of the Edwards aguifer, central Texas. Carbonates Evaporites 28:67–73.
- US Environmental Protection Agency (2018) Sole source aquifers for drinking water.
 Available at https://www.epa.gov/dwssa. Accessed December 18, 2018.
- Lindgren RJ (2004) Conceptualization and simulation of the Edwards Aquifer, San Antonio region, Texas. US Geological Survey Scientific Investigations Report 2004-5277 (US Geological Survey, Reston, VA), pp 1–154.
- Anaya R, Jones I (2009) Groundwater availability model for the Edwards-Trinity (Plateau) and Pecos Valley Aquifers of Texas. Texas Water Development Board Report 373 (Texas Water Development Board, Austin, TX), pp 1–115.
- Jones IC (2006) Defining groundwater flow characteristics in the northern segment of the Edwards Aquifer based on groundwater chemistry. Austin Geol Soc Bull 2:1–22.
- Maclay RW, Small TA (1983) Hydrostratigraphic subdivisions and fault barriers of the Edwards aquifer, south-central Texas, USA. J Hydrol (Amst) 61:127–146.
- Abbott PL (1975) On the hydrology of the Edwards Limestone, south-central Texas. J Hydrol (Amst) 24:251–269.
- Senger RK, Collins EW, Kreitler CW (1990) Hydrogeology of the northern segment of the Edwards Aquifer, Austin region. Bureau of Economic Geology Report of Investigations No. 192 (Bureau of Economic Geology, Austin, TX), pp 1–58..
- Smith BA, et al. (2014) Hydrologic influences of the Blanco River on the Trinity and Edwards Aquifers, central Texas, USA. Hydrogeological and Environmental Investigations in Karst Systems, eds Andreo B, Carrasco F, Durán JJ, Jiménez P (Springer, Berlin), pp 153–161.
- Smith BA, Hunt BB, Johnson SB (2012) Revisiting the hydrologic divide between the San Antonio and Barton springs segments of the Edwards aquifer: Insights from recent studies. Gulf Coast Assoc Geol Soc J 1:55–68.
- Fratesi SB, et al. (2015) Development of a finite-element method groundwater flow model for the Edwards Aquifer. SWRI project no. 20-17344–Final report prepared for the Edwards Aquifer Authority (Southwest Research Institute, San Antonio, TX), pp 1–193.
- Carmelli D, Cavalli-Sforza LL (1976) Some models of population structure and evolution. Theor Popul Biol 9:329–359.
- 95. Slatkin M, Voelm L (1991) F_{ST} in a hierarchical island model. Genetics 127:627–629.
- Pritchard JK, Stephens M, Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* 155:945–959.
- Chifman J, Kubatko L (2014) Quartet inference from SNP data under the coalescent model. Bioinformatics 30:3317–3324.
- 98. Yang Z, Rannala B (2014) Unguided species delimitation using DNA sequence data from multiple loci. *Mol Biol Evol* 31:3125–3135.

123. Säterberg T, Sellman S, Ebenman B (2013) High frequency of functional extinctions in ecological networks. *Nature* 499:468–470.
 124. Green RT, Painter SL, Sun A, Worthington SRH (2006) Groundwater contamination in

99. Rannala B, Yang Z (2017) Efficient Bayesian species tree inference under the mul-

100. Woodruff CM, Jr, et al. (1985) Edwards Aquifer—Northern segment, Travis, Williamson,

101. Hedin M (2015) High-stakes species delimitation in eyeless cave spiders (Cicurina,

102. Russell WH (1993) The Buttercup Creek karst, Travis and Williamson Counties, Texas:

103. Mike Warton and Associates (1997) Final report of findings: A study of cave habitats,

Geology, biology, and land development. Report prepared for the University Spe-

leological Society, a chapter of the National Speleological Society at The University

point recharge potential, and associated karst lands of the Buttercup Creek develop-

ment properties, including Buttercup Creek section 4 and phase v, Cedar Park, Wil-

Austin, Texas, USA. 13th Sinkhole Conference, National Cave and Karst Research Institute

in the Edwards Aguifer, northeastern Bexar and southern Comal Counties, Texas, US

Geological Survey Investigations Report 2007-5285 (US Geological Survey, Reston,

Springs based on hydrologic and geochemical data (2008-10), Bexar, Comal, and

Hays Counties, Texas. US Geological Survey Scientific Investigations Report 2012-

Symposium 2 (National Cave and Karst Research Institute, Carlsbad, NM), pp 353-365.

liamson County, Texas (Lumbermen's Investment Corporation, Austin, TX), pp 1-18.

104. Hauwert N, Cowan B (2013) Delineating source areas to cave drips and cave streams in

105. Otero CL (2008) Geologic, hydrologic, and geochemical identification of flow paths

106. Musgrove ML, Crow CL (2012) Origin and characteristics of discharge at San Marcos

107. de Queiroz K (2005) Ernst Mayr and the modern concept of species. Proc Natl Acad

108. Hey J (2006) Recent advances in assessing gene flow between diverging populations

109. US Fish and Wildlife Service (2009) Endangered and threatened wildlife and plants;

110. Lucas LK, Gompert Z, Ott JR, Nice CC (2009) Geographic and genetic isolation in

111. Smith HM, Potter FE (1946) A third neotenic salamander of the genus Eurycea from

112. Sweet SS (1984) Secondary contact and hybridization in the Texas cave salamanders

113. US Census Bureau (2017) Top 10 fastest-growing counties (percent change, counties

114. Jones IC (2003) Groundwater availability modeling: Northern segment of the Ed-

115. Hopkins J (1998) Areas experiencing significant ground-water decline, 1985-1995.

116. Ridgeway C, Petrini H (1999) Changes in groundwater conditions in the Edwards and

117. Mace RE, Chowdhury AH, Anaya R, Way S-C (2000) Groundwater availability of the

118. Bluntzer RL (1992) Evaluation of the ground-water resources of the Paleozoic and

119. Heitmuller FT. Reece BD (2006) Spatial data for Eurycea salamander habitats asso-

120. Zektser S, Loáiciga HA, Wolf JT (2004) Environmental impacts of groundwater

121. Perkin JS, et al. (2017) Groundwater declines are linked to changes in Great Plains

122. Bonacci O, Pipan T, Culver DC (2008) A framework for karst ecohydrology. Environ

stream fish assemblages. Proc Natl Acad Sci USA 114:7373-7378.

Board Report 339 (Texas Water Development Board, Austin, TX), pp 1-161.

with a population of 10,000 or more): July 1, 2015 to July 1, 2016. Available at https://

www.census.gov/content/dam/Census/newsroom/press-kits/2017/Top%2010%20Fastest-

wards Aquifer, Texas. Texas Water Development Board Report 358 (Texas Water

Texas Water Development Board Hydrologic Atlas No. 7 (Texas Water Development

Trinity Aquifers, 1987-1997, for portions of Bastrop, Bell, Burnet, Lee, Milam, Travis,

and Williamson Counties, Texas. Texas Water Development Board Report 350 (Texas

Trinity Aquifer, Hill Country area, Texas: Numerical simulations through 2050. Texas

Water Development Board Report 353 (Texas Water Development Board, Austin,

Cretaceous aguifers in the Hill Country of Central Texas. Texas Water Development

ciated with three aguifers in south-central Texas. US Geological Survey Data Series,

prepared in cooperation with the US Fish and Wildlife Service (US Geological Survey,

overdraft: Selected case studies in the southwestern United States. Environ Geol 47:

Eurycea neotenes and E. tridentifera. Copeia 1984:428-441.

partial 90-day finding on a petition to list 475 species in the southwestern United

States as threatened or endangered with critical habitat; proposed rule. Fed Regist

spring-associated Eurycea salamanders endemic to the Edwards Plateau region of

and Bell Counties (Austin Geological Society, Austin, TX), pp 1-110.

Dictynidae, Araneae) from central Texas, Mol Ecol 24:346-361.

of Texas at Austin (Univ of Texas, Austin), pp 1-82.

5126 (US Geological Survey, Reston, VA), p 94.

and species. Curr Opin Genet Dev 16:592-596.

Growing.pdf, p 1, Accessed December 18, 2018.

Water Development Board, Austin, TX), pp 1-42.

Development Board, Austin, TX), pp 1-83.

Board, Austin, TX), pp 1.

TX), p 122.

396-404

Reston, VA), pp 1-3.

Geol 56:891-900

Texas, Conserv Genet 10:1309-1319.

Texas, Herpetologica 3:105-109.

tispecies coalescent. Syst Biol 66:823-842.

VA), pp 1-57.

Sci USA 102:6600-6607.

- karst terranes. Water Air Soil Pollut Focus 6:157–170.
- Musgrove M, et al. (2016) Source, variability, and transformation of nitrate in a regional karst aquifer: Edwards aquifer, central Texas. Sci Total Environ 568:457–469.
- karst aquifer: Edwards aquifer, central Texas. *Sci Total Environ* 568:457–469.

 126. Texas Water Development Board (2017) State water plan. Available at www.twdb.
- texas.gov/waterplanning/swp/2017/index.asp. Accessed December 18, 2018.

 127. Banner JL. et al. (2010) Climate change impacts on Texas water: A white paper as-
- 127. Banner JL, et al. (2010) Climate change impacts on Texas water: A white paper assessment of the past, present and future and recommendations for action. Tex Water J 1:1–19.
- 128. McKay MG, McWilliams RH, eds (1993) Sierra Club v. Lujan, No. MO-91-CA-69, 1993 WL 151353, at *1 (W.D. Tex. Feb. 1).

- 129. Houston & Texas Central Railroad Co. v. East (1904) 98 Tex. 146, 81 S.W. 279.
- Wells S (2014) The "take" trifecta: Balancing the challenges of endangered species, private property interests, and state management authority in Texas water planning. Rev Litigat 33:429–468.
- 131. Vörösmarty CJ, Green P, Salisbury J, Lammers RB (2000) Global water resources: Vulnerability from climate change and population growth. *Science* 289:284–288.
- Tilman D, et al. (2001) Forecasting agriculturally driven global environmental change. Science 292:281–284.
- 133. Taylor RG, et al. (2012) Ground water and climate change. Nat Clim Chang 3:322–329.
- Famiglietti JS, Rodell M (2013) Environmental science. Water in the balance. Science 340:1300–1301.
- Mekonnen MM, Hoekstra AY, Hoekstra AY, Hoekstra AY (2016) Four billion people facing severe water scarcity. Sci Adv 2:e1500323.
- 136. Hayhoe K (2014) Climate change projections for the City of Austin. Draft Report (ATMOS Research and Consulting, Lubbock, TX), pp 1–9.
- Jiang X, Yang ZL (2012) Projected changes of temperature and precipitation in Texas from downscaled global climate models. Clim Res 53:229–244.
- 138. Stamm JF, et al. (2015) Historical and projected climate (1901–2050) and hydrologic response of karst aquifers, and species vulnerability in south-central Texas and western South Dakota. US Geological Survey Scientific Investigations Report 2014-5089 (US Geological Survey, Reston, VA), pp 1–74.
- Loáiciga HA, Maidment DR, Valdes JB (2000) Climate-change impacts in a regional karst aquifer, Texas, USA. J Hydrol (Amst) 227:173–194.
- 140. Peterson BK, Weber JN, Kay EH, Fisher HS, Hoekstra HE (2012) Double digest RAD-seq: An inexpensive method for de novo SNP discovery and genotyping in model and non-model species. PLoS One 7:e37135.
- 141. Catchen J, Hohenlohe PA, Bassham S, Amores A, Cresko WA (2013) Stacks: An analysis tool set for population genomics. *Mol Ecol* 22:3124–3140.
- 142. Eaton D, Overcast I (2018) ipyrad: Interactive assembly and analysis of RAD-seq data sets. Available at https://github.com/dereneaton/ipyrad. Accessed December 18, 2018.
- 143. Mastretta-Yanes A, et al. (2015) Restriction site-associated DNA sequencing, genotyping error estimation and de novo assembly optimization for population genetic inference. Mol Ecol Resour 15:28–41.
- 144. Ilut DC, Nydam ML, Hare MP (2014) Defining loci in restriction-based reduced representation genomic data from nonmodel species: Sources of bias and diagnostics for optimal clustering. *Biomed Res Int* 2014:675158.
- 145. Harvey MG, et al. (2015) Similarity thresholds used in DNA sequence assembly from short reads can reduce the comparability of population histories across species. *PeerJ* 3:e895.
- 146. Devitt T, et al. (2018) Restriction-site associated DNA sequences of Eurycea salamanders. NCBI SRA database. Available at https://www.ncbi.nlm.nih.gov/bioproject/?term=PRJNA504898. Deposited November 9, 2018.
- 147. Devitt T, et al. (2018) Data from "Species delimitation in endangered groundwater salamanders: Implications for aquifer management and biodiversity conservation." Dryad Digital Repository. Available at https://doi:10.5061/dryad.3k5c0sh. Deposited December 13, 2018.
- 148. Yang R-C (1998) Estimating hierarchical F-statistics. Evolution 52:950-956.
- 149. Goudet J, Raymond M, de Meeüs T, Rousset F (1996) Testing differentiation in diploid populations. *Genetics* 144:1933–1940.
- de Meeûs T, Goudet J (2007) A step-by-step tutorial to use HierFstat to analyse populations hierarchically structured at multiple levels. *Infect Genet Evol* 7:731–735.
- Goudet J (2005) HIERFSTAT, a package for R to compute and test hierarchical Fstatistics. Mol Ecol Notes 5:184–186.
- 152. R Core Team (2018) R: A language and environment for statistical computing (R Foundation for Statistical Computing, Vienna).
- Swofford DL (2002) PAUP*. Phylogenetic Analysis Using Parsimony (*and Other Methods) (Sinauer Associates, Sunderland, MA).
- 154. Huang H, He Q, Kubatko LS, Knowles LL (2010) Sources of error inherent in speciestree estimation: Impact of mutational and coalescent effects on accuracy and implications for choosing among different methods. Syst Biol 59:573–583.
- Reaz R, Bayzid MS, Rahman MS (2014) Accurate phylogenetic tree reconstruction from quartets: A heuristic approach. *PLoS One* 9:e104008.
- Fernández-Sánchez J, Casanellas M (2016) Invariant versus classical quartet inference when evolution is heterogeneous across sites and lineages. Syst Biol 65:280–291.
- Stamatakis A (2014) RAxML version 8: A tool for phylogenetic analysis and postanalysis of large phylogenies. *Bioinformatics* 30:1312–1313.
- Lewis PO (2001) A likelihood approach to estimating phylogeny from discrete morphological character data. Syst Biol 50:913–925.
- 159. Leaché AD, Banbury BL, Felsenstein J, de Oca AN-M, Stamatakis A (2015) Short tree, long tree, right tree, wrong tree: New acquisition bias corrections for inferring SNP phylogenies. Syst Biol 64:1032–1047.
- Sukumaran J, Holder MT (2010) DendroPy: A Python library for phylogenetic computing. *Bioinformatics* 26:1569–1571.
- 161. Miller MA, Pfeiffer W, Schwartz T (2010) Creating the CIPRES science gateway for inference of large phylogenetic trees. Proceedings of the Gateway Computing Environments Workshop (GCE-2010) (IEEE, Piscataway, NJ), pp 1–8.
- Rannala B, Yang Z (2003) Bayes estimation of species divergence times and ancestral population sizes using DNA sequences from multiple loci. Genetics 164:1645–1656.
- Rannala B, Yang Z (2013) Improved reversible jump algorithms for Bayesian species delimitation. Genetics 194:245–253.
- 164. Yang Z, Rannala B (2010) Bayesian species delimitation using multilocus sequence data. Proc Natl Acad Sci USA 107:9264–9269.
- 165. Yang Z (2015) The BPP program for species tree estimation and species delimitation. Curr Zool 61:854–865.